

TECHNICAL DATA SHEET

FITC Anti-Mouse CD62L (L-Selectin) (MEL-14)

Catalog Number: 35-0621

PRODUCT INFORMATION

Contents: FITC Anti-Mouse CD62L (L-Selectin) (MEL-14)

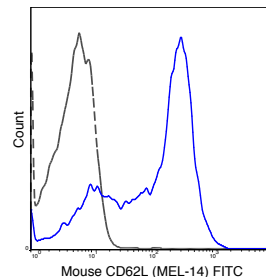
Isotype: Rat IgG2a, kappa

Concentration: 0.5 mg/mL

Clone: MEL-14

Reactivity: Mouse

Formulation: 10 mM NaH₂PO₄, 150 mM NaCl, 0.09% NaN₃,
0.1% gelatin, pH7.2



C57Bl/6 splenocytes were stained with 0.25 ug Anti-Mouse CD62L FITC (35-0621) (solid line) or 0.25 ug Rat IgG2a FITC isotype control (dashed line).

DESCRIPTION

The MEL-14 antibody is specific for mouse CD62L, also known as L-Selectin, a cell adhesion molecule which facilitates lymphocyte rolling on activated vascular endothelium and homing to high endothelial venules (HEV) as immune cells transmigrate from blood into peripheral tissues. L-Selectin is a member of a family of Selectin molecules which act together with the integrin family of adhesion molecules to mediate leukocyte-endothelial interactions. L-Selectin is characteristically expressed by neutrophils, and is also found on B cells, monocytes, granulocytes, and at varying levels on naive, effector and memory T cells. It is rapidly shed upon cell activation, releasing into the circulation a soluble form whose biological role is of particular interest in cancer biology research. The MEL-14 antibody may be used as a phenotypic marker for CD62L expression on a variety of immune cell types. Please note that CD62L (L-Selectin) itself is also referred to as MEL-14 in the literature.

PREPARATION & STORAGE

This monoclonal antibody was purified from tissue culture supernatant via affinity chromatography. The purified antibody was conjugated under optimal conditions, with unreacted dye removed from the preparation. It is recommended to store the product undiluted at 4°C, and protected from prolonged exposure to light. Do not freeze.

APPLICATION NOTES

This antibody preparation has been quality-tested for flow cytometry using mouse spleen cells, or an appropriate cell type (where indicated). The amount of antibody required for optimal staining of a cell sample should be determined empirically in your system.

REFERENCES

Lee L-F, Logronio K, Tu GH, Zhai W, Ni I, Mei L, Dilley J, Yu J, et al. 2012. Proc. Natl. Acad. Sci. 10.1073. (Flow cytometry). Harp JR, Gilchrist MA, and Onami TM. 2010. J. Immunol. 185:5751-5761. (in vivo blocking). Furukawa Y, Umemoto E, Jang MH, Tohya K, Miyasaka M, and Hirata T. 2008. J. Biol. Chem. 283: 12112-12119. (Immunoelectron microscopy). Li Y, Brazzell J, Herrera A, and Walcheck B. 2006. 108: 2275-2279. (Immunoprecipitation). Ochando JC, Yopp AC, Yng Y, Garin A, Li Y, Boros P, Llodra J, Ding Y, Lira SA, Krieger NR, and Bromberg JS. 2005. J. Immunol. 174: 6993-7005. (in vivo blocking, Flow cytometry). Zhao L-C, Shey M, Farnsworth M, and Dailey MO. 2001. J. Biol. Chem. 276: 30631-30640. (Immunoprecipitation). Suzuki A, Andrew DP, Gonzalo JA, Fukumoto M, Spellberg J, Hashiyama M, Takimoto H, Gerwin N, Webb I, Molineux G, Amakawa R, Tada Y, Wakeham A, Brown J, McNiece I, Ley K, Butcher EC, Suda T, Gutierrez-Ramos JC, and Mak TK. 1996. Blood. 87:3550-3562. (Stamper-Woodruff assay; in vivo blocking). Reichert RA, Jerabek L, Gallatin WM, Butcher EC, and Weissman IL. 1986. 136(10): 3535-3542. (Immunohistochemistry).