

TECHNICAL DATA SHEET

PE-Cy7 Anti-Mouse IFN gamma (XMG1.2)

Catalog Number: 60-7311

PRODUCT INFORMATION

Contents: PE-Cy7 Anti-Mouse IFN gamma (XMG1.2)

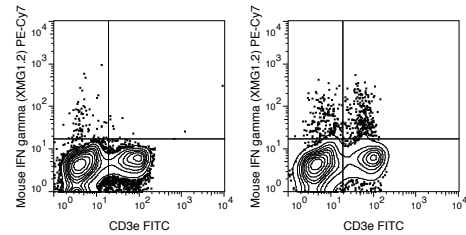
Isotype: Rat IgG1, kappa

Concentration: 0.2 mg/mL

Clone: XMG1.2

Reactivity: Mouse

Formulation: 10 mM NaH₂PO₄, 150 mM NaCl, 0.09% Na₃N, 0.1% gelatin, pH7.2



C57Bl/6 splenocytes were stimulated with PMA and Ionomycin (right panel) or unstimulated (left panel) and then stained with FITC Anti-Mouse CD3e (35-0031), followed by intracellular staining with 0.125 ug PE-Cy7 Anti-Mouse IFN gamma (60-7311).

DESCRIPTION

The XMG1.2 antibody is specific for mouse Interferon-gamma (IFN-g), a 20 kDa type II cytokine known for its central roles in protection against bacterial or viral pathogens and for its anti-tumor properties. IFN-g is secreted by several types of immune cells, which allow the cytokine to modulate innate immunity, when secreted by NK and NKT cells, and to function in support of adaptive immunity when secreted by Th1 and CD8+ T cells (CTLs). The XMG1.2 antibody is suitable for detection of intracellular IFN-g protein, e.g. by flow cytometry, as well as for quantitative analysis of the secreted protein by ELISA, when paired with an appropriate secondary antibody. This clone is also widely used for neutralization of the functional activity of IFN-g in a variety of assays.

PREPARATION & STORAGE

This monoclonal antibody was purified from tissue culture supernatant via affinity chromatography. The purified antibody was conjugated under optimal conditions, with unreacted dye removed from the preparation. It is recommended to store the product undiluted at 4°C, and protected from prolonged exposure to light. Do not freeze.

APPLICATION NOTES

This antibody preparation has been quality-tested for flow cytometry using mouse spleen cells, or an appropriate cell type (where indicated). The amount of antibody required for optimal staining of a cell sample should be determined empirically in your system.

REFERENCES

Choudhry N, Petry F, van Rooijen N, and McDonald V. 2012. *J. of Infect. Disease.* 206: 117-124. (in vivo neutralization) Cobb D and Smeltz RB. 2012. *J. Immunol.* 188: 3766-3773. (in vitro neutralization) Brown DM, Lee S, Garcia-Hernandez M, and Swain SL. 2012. *J. Virol.* 86: 6792-6803. (ELISA - detection) Yu H, Karunakaran KP, Jiang X, Shen C, Andersen P, and Brunham RC. 2012. *Infect. Immun.* 80: 1510-1518. (ELISA - detection) Kwon M-J, Ma J, Ding Y, Wang R, and Sun Z. 2012. *J. Immunol.* 188: 5887-5897. (in vitro induction of Th2 polarization) Barr TA, Shen P, Brown S, Lampropoulou V, Roch T, Lawrie S, Fan B, O'Connor RA, Anderton SM, Bar-Or Am Fillatreau S, and Gray D. 2012. *J. Exp. Med.* 209: 1001-1010. (flow cytometry) Cardona AE, Restrepo BI, Jaramillo JM, and Teale JM. 1999. *J. Immunol.* 162: 995-1002. (immunohistochemistry – frozen tissue) Kupfer A, Mosmann TR, and Kupfer H. 1991. *Proc. Natl. Acad. Sci.* 88: 775-779. (immunofluorescence microscopy)

NOTE: Please choose the appropriate format for each application. Citations are provided as a convenience to you; please consult Materials and Methods sections for additional details about the use of any product in these publications.

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